Protein fucosylation regulates synapsin Ia/Id expression and neuronal morphology in primary hippocampal neurons


*Howard Hughes Medical Institute and Division of Chemistry and Chemical Engineering, California Institute of Technology, 1200 East California Boulevard, Pasadena, CA 91125; †Ensemble Discovery Corporation, Cambridge, MA 02139; and ‡Department of Psychiatry, New York University School of Medicine/Nathan Kline Institute for Psychiatric Research, Orangeburg, NY 10962

Edited by Pietro De Camilli, Boyer Center for Molecular Medicine, New Haven, CT, and approved November 15, 2005 (received for review April 24, 2005)

Although fucose-(1-2)-galactose [Fuc(1-2)Gal] carbohydrates have been implicated in cognitive processes such as long-term memory, the molecular mechanisms by which these sugars influence neuronal communication are not well understood. Here, we present molecular insights into the functions of Fuc(1-2)Gal sugars, demonstrating that they play a role in the regulation of synaptic proteins and neuronal morphology. We show that synapsins Ia and Ib, synapse-specific proteins involved in neurotransmitter release and synaptogenesis, are the major Fuc(1-2)Gal glycoproteins in mature cultured neurons and the adult rat hippocampus. Fucosylation has profound effects on the expression and turnover of synapsin in cells and protects synapsin from degradation by the calcium-activated protease calpain. Our studies suggest that defucosylation of synapsin has critical consequences for neuronal growth and morphology, leading to stunted neurite outgrowth and delayed synapse formation. We also demonstrate that Fuc(1-2)Gal carbohydrates are not limited to synapsin but are found on additional glycoproteins involved in modulating neuronal architecture. Together, our studies identify important roles for Fuc(1-2)Gal sugars in the regulation of neuronal proteins and morphological changes that may underlie synaptic plasticity.

Materials and Methods

Neuronal Cultures and Immunocytochemistry. Hippocampal and cortical neurons were cultured and immunostained as described (12). Synaptin I knockout mice (13) were generously provided by H. T. Kao and P. Greengard (The Rockefeller University, New York). Antibody A46-B/B10 (5) was a generous gift from U. Karsten (Max-Delbrück Centre for Molecular Medicine, Berlin-Buch, Germany) and was incubated in 3% BSA (2.5 μg/ml) overnight at 4°C. The anti-tubulin (1:500; Sigma), anti-synapsin (1:5,000; Molecular Probes), and anti-spinophilin [1:10,000 (14)] antibodies were added in 3% BSA for 2 h at 37°C. Goat anti-mouse IgM-AlexaFlour 488 or goat anti-rabbit IgG AlexaFlour 568 (1:250; Molecular Probes) was added for 1 h at 37°C in 3% BSA.

Synaptic Vesicle Purification from Adult Rat Brain. Synaptic vesicles were purified by using a sucrose density gradient, as described (15).

Treatment of Cells with Deoxy-Galactose Analogues. Rat neuronal cultures were treated after 7 days, as described (12). Neuronal cultures from C57BL/6 embryonic day (E)16 mice were treated after 7 days in vitro (DIV) with 2-deoxygalactose (2-dGal) (0, 5, 10, or 15 mM) in PBS for 5 days. Neurons from C57BL/6 and synaptin I knockout postnatal day 0 mice were cultured for 2 days and then treated for 3 days with 15 mM 2-dGal. HeLa cells were seeded at 6 × 10⁵ cells per 60-mm dish in DMEM supplemented with 10% FCS and incubated at 37°C/5% CO₂ for 24 h. After pretreatment with the deoxy-galactose analogues (0.5–10 mM) for 1 h, the cells were transfected at ~60% confluence with the plasmid pCMV-FLAG-Synapsin Ia (see Supporting Text, which is published as supporting information online).

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Abbreviations: Fuc(1-2)Gal, fucose-(1-2)-galactose; LTP, long-term potentiation; DIV, days in vitro; En, embryonic day n; NSF, N-ethylmaleimide-sensitive factor; 2-dGal, 2-deoxy-α-galactose; 6-dGal, 6-deoxy-α-galactose; 6-deoxy-α-galactose.

§To whom correspondence should be addressed. E-mail: lhw@caltech.edu.

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information on the PNAS web site) and pSV-β-galactosidase (Promega) by using Lipopectamine 2000 (Invitrogen). After 22 h, the cells were harvested, resuspended in PBS, and either lysed in 1% boiling SDS (70% of the cells) or analyzed for transfection efficiency by using a β-galactosidase assay (30% of the cells). For synapsin degradation experiments, cells were treated with 2-dGal or PBS as above, followed by treatment 4 h posttransfection with bafilomycin A1 (100 nM, Acros, Geel, Belgium), MG132 (5 μM, Sigma), ammonium chloride (25 mM, Fisher), calpain inhibitor peptide (33 μM, Sigma), or calpeptin (2 μM, Alexis Biochemicals, Lausen, Switzerland). Cells were lysed as above after 15 h of treatment, resolved by SDS/PAGE, and analyzed by immunoblotting with chemiluminescence detection (Pierce).

Results
Expression of Fuc(1-2)Gal on Glycoproteins in the Hippocampus. We investigated whether Fuc(1-2)Gal glycoproteins are present in the hippocampus, a brain structure important for spatial learning and memory (16). Cell lysates from adult rat hippocampus, E18 hippocampus, and cultured embryonic hippocampal neurons were analyzed by Western blotting by using an antibody (A46-B/B10) selective for Fuc(1-2)Gal (5). Antibody A46-B/B10 has been shown to induce amnesia in animals (6), suggesting that it recognizes one or more physiologically relevant epitopes. We found that the Fuc(1-2)Gal epitope is present on several distinct proteins during neuronal development (Fig. 1A). In E18 hippocampal tissue, three major glycoproteins of ~55, 60, and 65 kDa are prominently observed, whose expression is significantly reduced in the adult hippocampus. In contrast, glycoproteins of ~73 and 75 kDa are found in mature cultured neurons and in adult brain tissue. Interestingly, expression of Fuc(1-2)Gal is observed on multiple proteins in developing neurons cultured for 4 and 7 DIV, periods when axons, dendrites, and functional synapses are being formed. Thus, expression and/or fucosylation of Fuc(1-2)Gal glycoproteins appear to change extensively during the course of neuronal development.

Fuc(1-2)Gal Is Enriched at Synapses. We next investigated the subcellular localization of Fuc(1-2)Gal sugars in neurons. Hippocampal neurons were cultured for 14 DIV to allow for synapse formation and were subsequently fixed, permeabilized, and coimmunostained with antibody A46-B/B10 and an antibody against the neuronal marker tubulin. The Fuc(1-2)Gal epitope exhibited a punctate pattern consistent with enriched localization to neuronal synapses (Fig. 1B). To examine whether the sugar was present at pre- or postsynaptic terminals, neurons were coimmunostained for Fuc(1-2)Gal and the presynaptic marker synapsin I or the postsynaptic marker spinophilin. Fuc(1-2)Gal labeling was observed in a subpopulation of the synapses (58 ± 2%; n = 350), overlapping with synapsin-positive puncta (Fig. 1C) and generally apposing spinophilin-positive puncta (Fig. 1D). Membrane delipidation by using methanol/chloroform did not alter the immunostaining pattern, which confirms the staining of glycoproteins rather than glycolipids (Fig. 6, which is published as supporting information on the PNAS web site). These findings indicate that Fuc(1-2)Gal sugars are enriched on glycoproteins in presynaptic nerve terminals.

Synapsins Ia and Ib Are the Major Fuc(1-2)Gal Glycoproteins in the Hippocampus. We next sought to identify neuronal glycoproteins modified by the Fuc(1-2)Gal epitope. Attempts to purify Fuc(1-2)Gal glycoproteins from brain extracts by using antibody A46-B/B10 were unsuccessful due to the relatively weak binding affinity of the antibody for the carbohydrate epitope. Fucose-specific lectins such as Lotus tetragonolobus lectin (LTL) and Ulex europaeus agglutinin I (UEA-I) also displayed weak affinity or broad specificity, resulting in limited enrichment of Fuc(1-2)Gal glycoproteins. To circumvent these challenges, we identified potential glycoproteins using a combination of subcellular fractionation, gel electrophoresis, and MALDI-TOF MS. Adult rat hippocampal lysates were enriched in synaptic proteins by using standard subcellular fractionation procedures. The crude synaptosomal fractions were resolved by 1D or 2D gel electrophoresis and analyzed by MALDI-TOF MS. Analysis revealed three potential Fuc(1-2)Gal-containing glycoproteins: synapsin Ia, synapsin Ib, and NSF. Eleven measured matching peptides covered 11.2% of the amino acid sequence (Table 1, which is published as supporting information on the PNAS web site). For NSF, 24 matching peptides were detected within 50 ppm accuracy, and the unmodified matching peptides covered 11.2% of the amino acid sequence (Table 1, which is published as supporting information on the PNAS web site).

To establish whether synapsins Ia/ib and NSF were indeed recognized by antibody A46-B/B10, each protein was immunoprecipitated and examined by Western blotting with antibody A46-B/B10. Upon immunoprecipitation, synapsins Ia and Ib were specifically detected by the antibody, whereas NSF was not recognized (Fig. 24). Deletion mutagenesis of synapsin Ia revealed that the synapsin fucosylation sites are localized to the D domain, a region...
found only in the synapsin I isoforms (Fig. 7, which is published as supporting information on the PNAS web site). In addition, loss of the fucosylated bands corresponding to synapsins Ia and Ib was observed by Western blot analysis of adult hippocampal lysates from synapsin I-deficient mice, confirming that synapsins Ia and Ib are the predominant Fuc(1-2)Gal glycoproteins (Fig. 8, which is published as supporting information on the PNAS web site). Treatment of purified synapsin I with PNGaseF and EndoH, enzymes that cleave N-linked oligosaccharides from proteins, did not abolish the interaction with antibody A46-B/B10, which suggests that the Fuc(1-2)Gal moiety is present on an O-linked glycan (data not shown). Together, these studies suggest that synapsins Ia and Ib are glycosylated in the D domain with an O-linked fucosyl oligosaccharide.

Characterization of the Carbohydrate Structure on Synapsin. Having identified the synapsins, we turned our attention to establishing the presence of the Fuc(1-2)Gal epitope. To gain insight into the structural determinants recognized by antibody A46-B/B10, we performed competition ELISA experiments using a series of fucose analogues (Fig. 2 B and C). Synapsins Ia and Ib from bovine brain were immobilized on a microtiter plate, and binding of antibody A46-B/B10 to synapsin was assessed in the presence of varying concentrations of the competitors. The monosaccharides ethyl α/β-L-fucopyranoside (L-Fuc-α-OEt and L-Fuc-β-OEt) and ethyl α/β-D-fucopyranoside (D-Fuc-α-OEt and D-Fuc-β-OEt), were synthesized from L- or D-fucose by using a modified Fischer glycosylation reaction (17). Disaccharide L-Fuc(1-2)Gal-β-OEt was synthesized from the known O-(2,3,4-tri-O-benzyl-α-L-fucopyranosyl)trichloroacetimidate (Fig. 9, which is published as supporting information on the PNAS web site), L-Fucose, L-Fuc-α-OEt, L-Fuc-β-OEt, and L-Fuc(1-2)Gal-β-OEt were effective at inhibiting antibody binding to synapsins Ia and Ib. Interestingly, the IC50 values of L-fucose, L-Fuc-α-OEt, and L-Fuc-β-OEt were comparable to that of the disaccharide, consistent with the specificity observed for some fucose-binding lectins such as Anguilla anguilla agglutinin (18). In contrast, the corresponding D-fucose sugars and naturally occurring sugars such as D-glucose (D-Glc), D-galactose (D-Gal), and N-acetyl-D-glucosamine (D-GlcNAc) did not compete with synapsin for antibody binding (Fig. 2C). These results suggest that antibody A46-B/B10 selectively recognizes L-fucose sugars on synapsin.

As independent confirmation, we examined the ability of L-fucose-specific lectins to bind to synapsin. Lotus tetragonolobus lectin (LTL) and Ulex europeus agglutinin I (UEA-I) lectins have been reported to interact strongly with terminal Fuc(1-2)Gal carbohydrates, whereas Anguilla anguilla agglutinin (AAA) prefers Fuc(1-3)Gal carbohydrates and interacts only weakly with Fuc(1-2)Gal (19). Consistent with the presence of a Fuc(1-2)Gal moiety on synapsin, both LTL and UEA-I readily detected synapsins Ia and Ib (Fig. 10, which is published as supporting information on the PNAS web site). However, AAA also recognized synapsin, indicating that lectins cannot be used to determine the nature of the fucose-galactose linkage on synapsin.

Because fucosidases have been shown to hydrolyze specific glycosidic linkages, we treated synapsins Ia and Ib with an α-(1-2)-fucosidase or an α-(1-3,4)-fucosidase from Xanthomonas manihotis. Rapid deglycosylation of synapsin was observed upon treatment with the α-(1-2)-fucosidase (Fig. 2D). In contrast, the α-(1-3,4)-fucosidase, which hydrolyzes both Fuc(1-3) and Fucα(1-4) linkages, had no effect on the fucosylation levels of synapsin, even after 6 h of treatment (Fig. 2D and data not shown). Together, these results provide strong evidence that synapsins Ia and Ib are covalently modified by the critical Fuc(1-2)Gal epitope.

Synapsin I Is Fucosylated in Various Subcellular Compartments. We next investigated the extent of fucosylation on neuronal synapsin I. Subcellular fractions of rat forebrain lysates were analyzed for levels of fucosylated or total synapsin (Fig. 3). Fucosylated synapsin was present in all subcellular fractions containing synapsin. Moreover,
the relative level of fucosylated synapsin to total synapsin was equivalent in the fractions examined. Quantitative analysis revealed that the membrane-associated to soluble ratio (LP2:LS2) of fucosylated synapsin was similar to that of synapsin (39:1 and 38:1 for fucosylated synapsin and synapsin, respectively; Fig. 11, which is published as supporting information on the PNAS web site). These results suggest that neuronal synapsin is extensively fucosylated in various subcellular compartments, and that fucosylation does not alter the subcellular localization of synapsin.

To gain insight into the stoichiometry of fucosylation, we compared synapsin I purified from bovine brain against a fucosylated BSA standard (Fig. 12, which is published as supporting information on the PNAS web site). Fucosylated [Fuc(1-2)Gal(1-4)GlcNAc] was conjugated to BSA by using reductive amination chemistry, and an epitope density of ≈3.0 ± 0.8 mol of fucose per BSA molecule was determined by using the Habeek assay (20). Comparison of the relative binding of antibody A46-B to synapsin I versus this standard revealed a stoichiometry of ≈1.5–3.2 Fucosyllactose (Fuc(1-2)Gal) epitopes per synapsin molecule. Together with the subcellular fractionation studies, these results indicate that a significant fraction of synapsin I is likely fucosylated in neurons.

**Inhibiting Synapsin Fucosylation Significantly Decreases Its Cellular Half-Life.** To investigate the impact of fucosylation on the functional properties of synapsin, we inhibited the fucosylation of synapsin in cells using 2-dGal. 2-dGal has been shown to prevent the fucosylation of glycoproteins (1). Upon cellular uptake, 2-dGal is converted via the Leloir pathway to the corresponding activated uridine diphosphate (UDP) analogue (1, 21). UDP-2-deoxy-galactose competes with UDP-galactose for incorporation into glycan chains and thereby terminates the chain by preventing formation of the Fucosyllactose linkage (1).

We first established that incubation of HeLa cells with 2-dGal leads to the biosynthesis of UDP-2-deoxy-galactose. Significant formation of UDP-2-deoxy-galactose was observed by LC-MS analysis of cell extracts, demonstrating that 2-dGal is an efficient unnatural substrate for the Leloir pathway enzymes (Fig. 13, which is published as supporting information on the PNAS web site). We next investigated the effects of 2-dGal on synapsin I expressed in HeLa cells. Cell lysates containing equivalent amounts of transfected protein were resolved by SDS/PAGE, and the fucosylation and protein levels of synapsin were measured by immunoblotting. Consistent with the presence of a Fucosyllactose epitope on synapsin, 2-dGal had a dramatic effect on the fucosylation level of synapsin (Fig. 4A). Unexpectedly, the 2-dGal treatment also led to a significant decrease in the level of synapsin protein. The effects appear to be specific to 2-dGal, because treatment with other deoxy-galactose sugars, including 6-deoxy-d-galactose (6-dGal), had no effect on either the fucosylation or protein levels of synapsin. Thus, 2-dGal was found to affect synapsin levels and fucosylation specifically on the C2 position of galactose.

Based on these results, we postulated that fucosylation might be critical for the half-life and turnover of synapsin in cells. We conducted pulse–chase experiments of synapsin Ia expressed in HeLa cells in the presence or absence of 2-dGal. Cells were pulse-labeled with [35S]-l-cysteine and [35S]-l-methionine, as described in Supporting Text, and chased for the indicated times. [35S]-labeled synapsin levels were measured by autoradiography. (C) Inhibition of the calcium-activated protease calpain using a calpain inhibitor peptide or calpeptin rescues synapsin from degradation. The lysosomal inhibitors bafilomycin A1 and ammonium chloride and the proteasome inhibitor MG132 had no effect on synapsin degradation. Synapsin I-transfected HeLa cells were treated with 2-dGal or a vehicle control (PBS), pulse-labeled with [35S]-l-cysteine and [35S]-l-methionine, as described in Supporting Text, and chased for the indicated times. [35S]-labeled synapsin levels were measured by autoradiography. (C) Inhibition of the calcium-activated protease calpain using a calpain inhibitor peptide or calpeptin rescues synapsin from degradation. The lysosomal inhibitors bafilomycin A1 and ammonium chloride and the proteasome inhibitor MG132 had no effect on synapsin degradation. Synapsin I-transfected HeLa cells were treated with the indicated inhibitors or vehicle control (DMSO, PBS, or MeOH) in the presence or absence of 2-dGal for 15 h. After lysis, the cell extracts were analyzed for synapsin expression levels by immunoblotting.

**Synapsin Degradation Is Mediated by the Calcium-Dependent Protease Calpain.** To investigate the molecular mechanisms responsible for synapsin turnover, cells expressing synapsin were treated with various inhibitors of protein degradation in the presence or absence of 2-dGal. Specifically, we used the lysosomal inhibitors bafilomycin A1 and ammonium chloride, the proteasome inhibitor MG132, and two inhibitors of the calcium-dependent protease calpain. With the exception of MG132, the inhibitors had minimal effects on synapsin expression levels in the absence of 2-dGal (Fig. 4C). As before, 2-dGal treatment of the cells significantly reduced levels of synapsin expression. Notably, inhibition of the protease calpain using a calpain inhibitor peptide or calpeptin rescued the effects of 2-dGal.
significantly attenuating the loss of synapsin, whereas the lysosomal and proteasomal inhibitors could not rescue synapsin from degradation. These data suggest that fucosylation protects synapsin from rapid degradation mediated at least in part by the Ca²⁺-dependent protease calpain.

**Fucosylation Modulates the Expression of Synapsin in Neurons and Neurite Outgrowth.** To examine the effects of 2-dGal on synapsin fucosylation in neurons, neurons were cultured for 7 DIV to allow for adequate expression of synapsin and subsequently treated with either 2- or 6-dGal. 2-dGal dramatically reduced the expression of synapsin I in cultured neurons (Fig. 5A). Importantly, the effects of 2-dGal appeared to be selective, because treatment with another deoxy sugar, 6-dGal, did not alter the expression of synapsin. Moreover, the effects of 2-dGal were specific to synapsin as the expression of other synaptic proteins, including NSF, synaptotagmin, syntaxin, PSD-95, the AMPA receptor subunit GluR1, and spinophilin, was unchanged by the 2-dGal treatment.

Because the synapsins play important roles in neuronal development and synaptogenesis (13, 23), we investigated whether 2-dGal might influence neuronal growth and morphology. Hippocampal neurons were cultured for 7 DIV as above to establish synapses and subsequently incubated for 3–5 days with 2-dGal at various concentrations. Treatment with 2-dGal induced a dramatic retraction of neurites and collapse of synapses, whereas 6-dGal had no effect (Fig. 5B and Fig. 14, which is published as supporting information on the PNAS web site). The effects of 2-dGal could be partially rescued by subsequent incubation of the neurons with D-Gal, which is expected to reestablish the Fuco(1-2)Gal linkage (Fig. 5B; 2.06 ± 0.14-fold rescue; n = 50, P < 0.0001). These results suggest that disruption of the Fuco(1-2)Gal linkage on neuronal glycoproteins has a profound impact on neurite outgrowth and neuronal morphology.

One potential mechanism by which 2-dGal might influence neuronal morphology is by regulating the function and/or expression of synapsin in presynaptic terminals. Notably, the phenotypic effects of 2-dGal on neurite outgrowth at 5 mM concentration are similar to deletion of the synapsin I gene, which results in retarded neurite outgrowth and delayed synapse formation (13) (Fig. 14). However, because other neuronal proteins bear the Fuco(1-2)Gal modification (Fig. 1A), these proteins might also contribute to the morphological effects observed upon fucosylation.

To examine the relative contribution of synapsin I to the effects of 2-dGal, neurons were cultured from synapsin I-deficient or wild-type mice for 2 days, treated in the presence or absence of 2-dGal for 3 days, and examined by confocal fluorescence microscopy. Fig. 5C, a and b are representative images of untreated wild-type and synapsin I-deficient neurons after 5 days in culture, respectively. Neurons from wild-type mice treated with 2-dGal had shorter neurites relative to their untreated wild-type counterparts (Fig. 5C, compare c and a). The effects of defucosylation at a concentration of 15 mM 2-dGal were more pronounced than elimination of the synapsin I gene (Fig. 5C, compare c and b). Finally, treatment with 2-dGal induced a more dramatic neurite retraction in wild-type relative to synapsin-deficient neurons (Fig. 5C, compare c and d). Although the length and extensive overlap among processes for untreated wild-type neurons precluded a quantitative analysis of neurite length, 2-dGal treatment led to neurite retraction and enabled quantification. We found that synapsin-deficient neurons displayed longer neurites than wild-type neurons upon treatment with 2-dGal (Fig. 5C, bar graph).

**Discussion**

Increasing evidence has linked synaptic activity with changes in the levels of protein fucosylation in the brain. For instance, both task-specific learning and LTP have been shown to enhance protein fucosylation at the synapse (7, 8). Moreover, the activity of fucosyltransferases increases substantially during synaptogenesis (10) and upon passive avoidance training in animals (11). Together, these studies suggest that protein fucosylation may relate to the dynamic regulation of synaptic proteins. Studies have implicated a particular carbohydrate, Fuco(1-2)Gal, in cognitive processes such as learning and memory (1–4). Although Fuco(1-2)Gal has been postulated to covalently modify synaptic glycoproteins, the identity of such proteins has remained elusive. In this study, we identify synapsins Ia and Ib as the major Fuco(1-2)Gal glycoproteins in maturing neuronal cultures and the adult rat hippocampus. Our results provide molecular insights into the functions of Fuco(1-2)Gal and demonstrate that these carbohydrates play a critical role in the regulation of synaptic proteins.

The synapsins are a family of highly conserved neuron-specific proteins that are associated with synaptic vesicles (24). Studies indicate that these proteins regulate multiple aspects of neuronal function. The synapsins have been shown to modulate neurotransmitter release by regulating the supply of releasable vesicles during periods of high activity (24, 25). In addition, synapsin I has recently been found to control synaptic vesicle dynamics in developing neurons via a cAMP-dependent pathway (26). Accordingly, synapsin-deficient mice show reduced numbers of synaptic vesicles within nerve terminals and exhibit significant alterations in neuro-
transmitter release and synaptic depression (24, 27). The synapsins have also been implicated in diverse aspects of neuronal development, including axon outgrowth, nerve terminal development, synapse formation, and synapse maintenance (13, 28, 29).

Our studies indicate that fucosylation of synapsin critically impacts its exocytosis and turnover in presynaptic nerve terminals. The potential to modulate the expression level of synapsin is expected to have important consequences for neuronal function. For instance, the addition of exogenous synapsin I to embryonic Xenopus spinal neurons has been shown to accelerate structural and functional maturation of neuromuscular synapses, including the early compartmentalization of synaptic vesicles into nerve terminals and a mature form of quantal secretion (30, 31). Conversely, reduction in synapsin expression levels also has profound effects: synapsin-deficient mice exhibit significant delays in axonal extension, neuronal differentiation, and synapse formation.

We found that defucosylation of synapsin promoted its degradation by calcium-activated calpain proteases, which are a family of nonlysosomal neutral cysteine proteases. These observations corroborate and extend recent findings suggesting a role for calpain in the regulation of both synaptic transmission and neuronal morphology (32, 33). Interestingly, calpain has been suggested to be critical in regulating depleted vesicle stores in sensory motor synapses of Aplysia via a mechanism involving the cleavage of potential substrates such as synapsin (34). In addition, calpain was found to degrade synaptosomal-associated protein of 25 kDa (SNAP-25), a protein essential for neurotransmitter release, in a calcium-dependent manner (35). The proteolytic activity of calpain was also shown to induce cytoskeletal rearrangements, leading to both facilitation and inhibition of neurite outgrowth (33, 36, 37). Although further studies will be necessary to understand more fully the mechanisms of calpain-mediated synapsin degradation, our results suggest an expanded role for this protease family in presynaptic nerve terminals and reveal that fucosylation has profound effects on the half-life of synapsin, possibly preventing synapsin from calcium-activated degradation.

In this study, we also investigated the role of synapsin fucosylation on neuronal growth and morphology. 2-dGal, a small molecule inhibitor of Fuco(1-2)Gal linkages, served as a valuable tool to defucosylate synapsin and dissect the role of the carbohydrate in modulating synapsin function. Treatment of neurons with 2-dGal led to stunted neurite outgrowth and delayed synapse formation. Moreover, significant differences were observed between wild-type and synapsin-deficient neurons upon treatment with 2-dGal. We believe that the extent of neurite retraction in synapsin-deficient mice is less pronounced because the primary target of 2-dGal, synapsin I, is missing. Indeed, the bar graph shown in Fig. 5C likely represents a lower estimate of the contribution of synapsin because neurites from synapsin-deficient neurons are shorter than those from wild-type neurons before treatment with 2-dGal. Based on these results, we propose that defucosylation may disrupt synapsin function, leading to its degradation and neurite retraction. Although further studies will be needed to resolve whether synapsin fucosylation stimulates or inhibits neurite outgrowth, these results strongly support the notion that synapsin fucosylation plays a role in modulating neuronal growth and morphology.

Our data also implicate other Fuco(1-2)Gal glycoproteins in the regulation of neuronal morphology. We demonstrate that Fuco(1-2)Gal carbohydrates are not limited to synapsin but are found on several additional proteins in developing neurons. Expression of the sugar and/or these glycoproteins changes dramatically during the course of neuronal development. We found that disruption of synapsin fucosylation contributed, but was not fully sufficient, to account for the striking neurite retraction induced by 2-dGal. For instance, 2-dGal had stronger effects on neurite outgrowth at high concentrations relative to deletion of the synapsin I gene, suggesting that 2-dGal may disrupt the fucosylation of other Fuco(1-2)Gal glycoproteins that influence neuronal morphology. Moreover, 2-dGal was still capable of inducing partial neurite retraction in synapsin-deficient neurons and young cultured neurons where synapsin expression is low (12). Thus, Fuco(1-2)Gal sugars appear to modulate the functions of multiple proteins involved in neuronal morphology and exert their effects via several distinct molecular mechanisms.

Finally, our findings may shed light on behavioral and electrophysiological studies implicating Fuco(1-2)Gal in long-term memory storage. Alterations in neuronal morphology, such as dynamic changes in dendritic spine number and shape, occur during memory consolidation and LTP (38, 39). Future studies will investigate whether Fuco(1-2)Gal sugars and their associated glycoproteins contribute to structural remodeling events that underlie synaptic plasticity.

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